

PANAMANIAN FOREST MAMMALS AS CARRIERS OF *SALMONELLA**

MIGUEL KOURANY, LARRY BOWDRE,[†] AND ARISTIDES HERRERA[‡]

Gorgas Memorial Laboratory, Apartado 6991, Panamá 5, Panamá

Abstract. Enteric bacteria pathogenic to man were sought in a total of 974 forest mammals collected from a variety of sites in rural and jungle areas of Panamá. The highest incidence of infection among the mammals was observed during the Panamanian dry season, which normally extends from January through April. A minimum of 10 *Salmonella* serotypes including, three of the *Arizona* group and *Edwardsiella tarda*, was isolated. Opossums of the genera *Philander*, 11 of 54 (20.1%), and *Didelphis*, 12 of 102 (11.8%) demonstrated high infection rates. One sloth of the genus *Choloepus* and specimens of two genera of rodents also were infected to varying degrees: 1 (11.1%) of 9 *Choloepus*, 8 (1.1%) of 704 *Proechimys* and 1 (16.7%) of 6 *Diplomys*.

Due to its topographic and climatic diversity, and despite its small geographic area (30,000 sq. miles), Panamá has a large mammalian fauna. As many as 214 mammals are known to occur on the Isthmus of Panamá.¹ Studies elsewhere have shown that a large number of species of mammals in many parts of the world are naturally infected by *Salmonella*.²⁻⁴ Previous surveys for enterobacterial pathogens among the wild animal populations in Panamá have recently been carried out to determine what role they might play as reservoirs for enteric pathogens. Those surveys were limited to nonhuman primates,⁵ various reptile and amphibian species,⁶ and elements of the rodent and marsupial fauna collected along a proposed site for a sea-level canal in eastern Panamá.⁷

The above studies revealed a variety of diarrhea-producing agents such as *Salmonella* and *Arizona* in a number of the more common species of animals found in this country. Such animals were without symptoms of disease, and perhaps may be the principle reservoir for human infections with *Salmonella* in remote rural and jungle communities.

Since rodents, opossums, and other mammals

occur throughout Panamá in high population densities, and a number of them proved to be harboring *Salmonella*, it was believed necessary to extend our studies to other geographic areas in this country. Thus, additional species would be included if we were to learn more about the variety and relative frequency of pathogenic enteric bacteria in wild mammals that find their best ecologic conditions in the vicinity of man.

The present investigation was initiated in March 1969, incidental to other studies, and continued through November 1971.

MATERIALS AND METHODS

Field Methods

Mammals were collected over a period of almost 3 years in all seasons and in a variety of habitats in different areas of the Republic of Panamá (Fig. 1 and Table 1). Most were trapped in the Pacific and Atlantic lowlands, and a few were collected along the Panama Canal. Small animals, such as rodents, were captured by setting Sherman traps. Larger animals such as opossums were caught in wire traps of various sizes (National Live Trap Corporation, P. O. Box 302, Tomahawk, Wisconsin 54487). The traps were baited with banana, corn, or toasted coconut. Sloths were captured by hand; bats were netted in free flight. All animals were taken to base camps in the field and kept in 25 × 25 × 35 cm stainless steel cages. Rodents were fed Lab-Blox (Allied Mills, 2205 Lamar Avenue, Memphis, Tenn. 38114). Opossums and other mammals were offered bananas, oranges, and yucca. The length of time they were

Accepted 8 November 1975.

* Address for reprint requests: Gorgas Memorial Laboratory, P. O. Box 2016, Balboa Heights, Panama Canal Zone.

[†] Present address: The Museum, Michigan State University, East Lansing, Michigan 48823.

[‡] Present address: Av. Elmer Faucett 474, Dpt. 205, Lima, Perú.

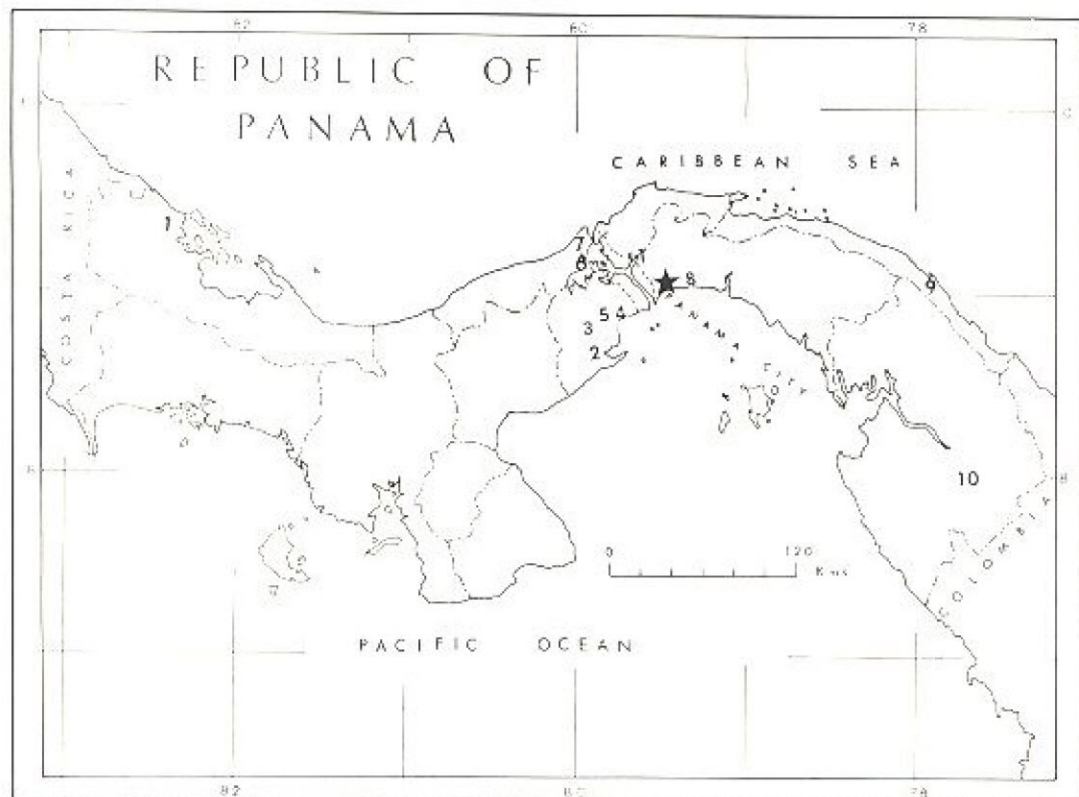


FIGURE 1. Map showing major collection sites in the Republic of Panamá. Locality No. 1, Almirante; No. 2, Cerro Campana; No. 3, Aguacate; No. 4, La Chorrera; No. 5, Santa Rita; No. 6, Achioté; No. 7, Las Tablitas; No. 8, Juan Díaz; No. 9, Sasardi; No. 10, Rio Mono; and No. 11, Madden Lake. Panama City is shown by the star.

held in the field never exceeded 1 week. None of the animals exhibited overt signs of disease on arrival at the Gorgas Memorial Laboratory in Panama City, where they were kept individually in cages for a variable length of time.

Laboratory Methods

Small animals (rodents and marsupials) were killed by chloroform, or by cardiac exsanguination after ether anesthesia. The large mammals (edentates and carnivores) after being tranquilized with Sernylan (Bio-Ceutic Laboratories Inc., Saint Joseph, Missouri 64502) were bled from the heart.

Aseptic techniques were used to remove portions of intestine and its contents from each animal immediately after death. The specimens were transferred with sterile swabs to selenite broth. Following incubation for 18 hours, aliquots

of the selenite broth were streaked on *Salmonella-Shigella* (SS) agar, MacConkey's (MC) agar, and bismuth sulfite (BS) agar plates. All plates were examined after incubation, according to the procedures recommended by Edwards and Ewing.⁸

Up to 10 nonlactose-fermenting colonies from each SS and MC plate were streaked on triple-sugar-iron (TSI) agar slants, and at least five suspected colonies were transferred from the BS plates to TSI agar and to lysine-iron-agar (LIA) slants. After overnight incubation, the cultures were studied by biochemical and serological methods described elsewhere.⁸ Examination was done only for *Salmonella*, *Shigella*, *Arizona*, and *Edwardsiella*. Following preliminary identification and serological grouping, cultures were submitted to the Enteric Bacteriology Unit, Center for Disease Control, Atlanta, Georgia for serotype identification.

TABLE 1
Panamanian forest mammals sampled for enterobacterial pathogens

Species	Number of specimens by locality											Total	
	Archife	Aguazote	Almirante	Campo	Chorro	Juan Diaz	Jas Talarias	Rio Mono	Santa Rita	Susudi	Madison Lake		Other
Marsupialia													
<i>Cuscomys derbianus</i>	-	-	-	-	-	-	-	-	8	-	-	1	6
<i>Marmosa rubicincta</i>	-	-	-	-	-	-	-	-	1	16	-	-	17
<i>Pithecheilus opossum</i>	2	5	-	3	-	-	-	-	42	-	2	-	54
<i>Metachirus nudicaudatus</i>	1	-	-	-	-	-	-	-	1	14	-	-	16
<i>Didelphis marsupialis</i>	7	-	-	3	4	-	1	-	85	-	1	1	102
Chiroptera													
<i>Pterostomus hastatus</i>	-	-	-	-	-	-	-	-	-	-	2	2	4
Edentata													
<i>Bradypus infuscatus</i>	-	-	-	-	-	-	10	-	-	-	-	2	12
<i>Choloepus hoffmanni</i>	-	8	-	-	-	-	-	-	-	-	-	1	9
<i>Dasylops novaeinactis</i>	-	-	-	-	-	-	-	-	-	-	-	1	1
Rodentia													
<i>Heteromys desmarestianus</i>	-	-	-	-	-	-	-	-	-	-	1	-	1
<i>Oryzomys coligiosus</i>	-	-	-	-	-	-	-	-	-	3	-	-	3
<i>Oryzomys cupido</i>	-	-	-	-	-	-	-	-	-	1	-	-	1
<i>Tytomys panamensis</i>	4	-	-	-	-	-	-	-	-	-	-	-	4
<i>Signodon hispidus</i>	-	-	-	-	-	-	-	-	6	-	-	-	6
<i>Rattus rattus</i>	-	-	-	-	3	-	-	-	7	-	1	-	11
<i>Proechimys semispinosus</i>	28	5	59	-	-	7	8	3	516	25	10	1	704
<i>Diplomys labilis</i>	-	-	-	-	-	-	-	-	6	-	-	-	6
Carnivora													
<i>Potos flavus</i>	-	-	-	-	-	-	-	-	-	2	-	-	2
<i>Bassaricyon gabbii</i>	-	-	-	-	-	-	-	-	-	1	-	-	1
Totals	42	18	113	6	7	7	19	5	665	62	17	9	974

TABLE 2
Incidence of enterobacterial pathogens in Panamanian forest mammals

Scientific name	Common name	No. examined	Positive	
			No.	%
Marsupialia				
<i>Caluromys derbianus</i>	Woolly opossum	6	0	—
<i>Marmosa robinsoni</i>	Brown murine opossum	17	0	—
<i>Philander opossum</i>	Four-eyed opossum	54	11	20.1
<i>Metachirus nudicaudatus</i>	Brown-masked opossum	16	0	—
<i>Didelphis marsupialis</i>	Common opossum	102	12	11.8
Total Marsupialia		195	23	11.8
Edentata				
<i>Bradypus infulcatus</i>	Three-toed sloth	12	0	—
<i>Choloepus hoffmanni</i>	Two-toed sloth	9	1	11.1
<i>Dasyurus novemcinctus</i>	Nine-banded armadillo	1	0	—
Total Edentata		22	1	4.5
Rodentia				
<i>Heteromys desmarestianus</i>	Spiny pocket mouse	1	0	—
<i>Oryzomys caliginosus</i>	Rice rat	3	0	—
<i>Oryzomys capito</i>	Rice rat	1	0	—
<i>Tylomys panamensis</i>	White-tailed rat	4	0	—
<i>Sigmodon hispidus</i>	Cotton rat	20	0	—
<i>Rattus rattus</i>	Rooi rat	11	0	—
<i>Proechimys semispinosus</i>	Spiny rat	704	8	1.1
<i>Diplomys labilis</i>	Arboreal spiny rat	6	1	16.7
Total Rodentia		750	9	1.2
Others				
<i>Bassaricyon gabbii</i>	Olingo	2	0	—
<i>Potos flavus</i>	Kinkajou	1	0	—
<i>Phyllostomus hastatus</i>	Spear-nosed bat	4	0	—
Total others		7	0	—
GRAND TOTALS		974	33	3.4

RESULTS

A total of 974 forest mammals were trapped, and represented five orders: Rodentia, Marsupialia, Edentata, Carnivora and Chiroptera (Table 1). A rodent (*Proechimys semispinosus*) and two marsupial species (*Philander opossum* and *Didelphis marsupialis*) comprised 88.5% of the sample, and 16 different species represented the remaining 11.5%. Other important species examined included sloths and carnivores, which were not as abundant or were not successfully trapped during our studies. The order Chiroptera was represented by four specimens of *Phyllostomus hastatus*.

The results of bacteriological cultures for enteric pathogens in these animals are summarized in Table 2; mainly *Salmonella* and *Arizona* were

recovered from 33 (3.4%) of the 974 animals examined. Organisms were demonstrated in the orders Marsupialia (11.8%), Edentata (4.5%) and Rodentia (1.2%). Among these orders, the incidence of infection varied considerably in relation with the species of animals studied and the season of the year. The highest infection rates (Table 2) were determined in *P. opossum* (20.1%) and *D. marsupialis* (11.8%), the two most common marsupials captured. The most numerous of the animals surveyed was the spiny rat, *P. semispinosus*, in which 8 (1.1%) of 704 specimens harbored enterobacterial pathogens. Infected *Proechimys* and *Diplomys* came from the Central Pacific side of Panamá.

The host distribution and frequency of the

TABLE 3
Frequency and host distribution of enterobacterial pathogens in Panamanian forest mammals

Bacterial species	Number of times strains isolated from host species					Total (%) strains isolated
	<i>Choloepus</i> <i>boljmanni</i> (1)*	<i>Didelphis</i> <i>marxipetris</i> (12)	<i>Diplomys</i> <i>labilis</i> (11)	<i>Philander</i> <i>opossum</i> (11)	<i>Proechimys</i> <i>semipinosus</i> (8)	
<i>S. newport</i>	—	3	1	5	2	11 (31.3)
<i>S. san diego</i>	—	4	—	1	—	5 (14.2)
<i>S. sandsvalli</i>	—	1†	—	—	2	3 (8.5)
<i>S. bonair</i>	—	1†	—	—	—	1 (2.9)
<i>S. oslo</i>	—	1	—	—	—	1 (2.9)
<i>S. miami</i>	—	—	—	1	—	1 (2.9)
<i>S. panama</i>	—	—	—	—	1	1 (2.9)
<i>S. wassenaar</i>	—	—	—	1	1	2 (5.7)
<i>S. rubislaw</i>	—	—	—	1‡	—	1 (2.9)
<i>Salmonella</i> , group II	—	1	—	—	1	2 (5.7)
<i>A. hinsavii</i> 16: 22, 21	—	—	—	—	1	1 (2.9)
<i>A. hinsavii</i> 26: 33, 31	1	—	—	—	—	1 (2.9)
<i>A. hinsavii</i> 30: 23, 25	—	1	—	—	—	1 (2.9)
<i>E. tarda</i>	—	1	—	3‡	—	4 (11.4)
TOTAL	1	13	1	12	8	35 (100.0)

* Number of animals found infected.

† One *Didelphis* with *S. bonair* and *S. sandsvalli*.

‡ One *Philander* with *S. rubislaw* and *E. tarda*.

bacterial species isolated are given in Table 3. In general, of the 35 strains recovered, *Salmonella newport* (31.4%), *S. san diego* (14.2%), and *E. tarda* (11.4%) were the most prevalent and accounted for 57% of all strains recovered. Ten *Salmonella* and three *Arizona* serotypes were represented, *Edwardsiella tarda*, whose relationship to diarrhea in humans is under investigation (Kourany, M., unpublished data), was isolated on four different occasions from opossum (*Didelphis* and *Philander*) hosts.

Nine of the 11 infected *Philander* were captured in central Panamá, in the dry season, while the other two were taken in the wet season. Two of these opossums harbored *Edwardsiella* alone while another was infected with both *Salmonella* and *Edwardsiella* (Table 3); the remaining eight harbored *Salmonella* only. Among the 102 *Didelphis* examined, 12 were infected with either *Salmonella*, *Arizona*, or *Edwardsiella*. The majority, however, harbored salmonellae (Table 3) and were collected in the dry season; only 3 of the infected 12 were caught in the wet season (Table 4). Opossums belonging to other genera (*Caluromys*, *Marmosa*, and *Metachirus*) were not found infected with pathogens.

Of the 750 rodents sampled, 9 (1.2%) were positive for enterobacterial pathogens. Among the

eight genera tested, infected rodents were found in only two (Table 2). Only one infected *Diplomys* and eight infected *Proechimys* from central Panamá were collected. The infected *Diplomys* rat was captured in the dry season; all but one of the *Proechimys* which harbored pathogens were trapped in the wet season (Table 4).

No enterobacterial pathogens were isolated from bats (*Phyllostomus*) or carnivores (*Bassaricyon* and *Potos*), but the number of specimens studied was small. Among the edentates examined, *Arizona* was isolated only from a single two-toed sloth (*Choloepus*) collected in central Panamá at El Aguacate in the dry season (Tables 2 and 4).

TABLE 4
Infection rates for enterobacterial pathogens in forest mammals according to season

Host mammal	Wet season			Dry season		
	No. examined	Positive No.	%	No. examined	Positive No.	%
<i>Philander</i>	17	2	11.8	37	9	24.3
<i>Didelphis</i>	45	3	6.7	57	9	15.8
<i>Choloepus</i>	6	0	—	3	1	—
<i>Proechimys</i>	472	7	1.5	232	1	0.4
<i>Diplomys</i>	4	0	—	2	1	—
TOTAL	544	12	2.2	331	21	6.3



FIGURE 2. Extensive deforestation in Panamá and the opening of large tracts of forest for cattle raising and agriculture is disturbing the ecological balance by limiting the natural cover and altering the food habits of many mammalian species. Notice hut (center) amidst what once was jungle.

DISCUSSION

This study indicated that diarrheal producing agents, such as *Salmonella*, are present among some elements of the wild mammalian fauna of Panamá and that they might serve as important reservoirs. *Salmonella* was the most prevalent of the pathogens isolated and at least 10 different serotypes were identified. *Arizona* as well as *Edwardsiella* was found, but was not as common as *Salmonella*, pointing to a diversity of harbored enteric bacterial species.

The mammals with the highest infection rates are common scavengers and are widely distributed in rural and jungle areas. They seem to thrive best in the vicinity of man, thus emphasizing their probable role as incidental reservoirs of *Salmonella* organisms. Since the distribution of these organisms in man is closely linked with that in the animal population of man's environment, these organisms are more frequently transmitted from animals to man than from man to man or from man to other animals.⁹ All of the serotypes listed in Table 3, except *S. bonair* and *S. wassenaar*, have been implicated in salmonellosis of man in rural or urban parts of Panamá.^{9, 10-12} Human infections with *E. tarda* and with *Arizona* organisms also have been detected, but the *Arizona* serotypes isolated were different from those found in the present study (Kourany, M., unpublished data).

The highest incidence of infection among the mammals was observed during the dry season,

which normally extends from January through April. Overall infection rates for the enterobacterial pathogens were about three times as high in animals trapped during the dry months (Table 4) than in those collected in the wet months ($p < 0.003 > 0.002$). This is particularly true for the two opossum species most commonly infected, *P. vossium* and *D. marsupialis*. It should be pointed out that these species are much more predatory and active during the dry season when they are constantly in search of food and this could account for the higher infection rates due to greater contact with sources of infection.

Most of the infected animals in the present study were caught in or near areas of human activity and could serve as a source of infection for man, his livestock, poultry, and other domestic animals. For example, the foraging habits of *D. marsupialis* bring this animal into closer contact with humans. This may pose an important public health problem since those species of animals capable of harboring enteric pathogens occur throughout Panamá in high densities and may contribute to health hazards in the rural areas.

With a high potential for social and economic growth, Panamá is developing at a fast pace. Roads and communication networks rapidly are being expanded through jungle and rural areas. In many regions, extensive deforestation is taking place. Large tracts of forest are being destroyed by traditional slash-and-burn methods (Fig. 2) as individual farms are opened for cattle raising and agriculture.

The chronically inadequate sanitation that prevails in these regions is most likely to be aggravated in the near future as large segments of the human population from central and western Panamá migrate to settle and colonize the newly opened lands. This situation is upsetting the ecological balance of the forest and is most likely to affect the stability of the wild mammalian fauna. Their natural cover and food habits may be affected and may bring man in closer contact with them, thus increasing many of the diseases, such as salmonellosis, that are transmitted from animals to man. Problems of potable water are compounded by a general lack of safe disposal of feces. The result is that animals harboring and shedding enterobacterial pathogens in close proximity to man, like the common and the four-eyed opossums, may conceivably be contributing fac-

tors in maintaining the endemicity of diarrheal disease of man in Panamá.

ACKNOWLEDGMENTS

The authors are deeply grateful to Dr. George J. Hermann of the Center for Disease Control, Atlanta, Georgia, for the determination of serotypes of *Salmonella* and *Arizona*.

REFERENCES

1. Handley, C. O., 1966. Checklist of the mammals of Panamá. Pages 753-795 in Wenzel, R. L., and Tipton, V. J. (eds.), *Ectoparasites of Panamá*. Field Museum of Natural History, Chicago.
2. Taylor, J., 1968. *Salmonella* in wild animals. *Symp. Zool. Soc. London*, No. 24: 51-73.
3. Van Oye, E., 1964. Les salmonelloses en Afrique Centrale. Pages 354-365 in Van Oye, E. (ed.), *The World Problem of Salmonellosis*. Dr. W. Junk Publ., The Hague.
4. Edwards, P. R., Bruner, D. W., and Moran, A. B., 1948. The genus *Salmonella*: Its occurrence and distribution in the United States. *Kentucky Agr. Exp. Sta. Bull.*, 525.
5. Kourany, M., and Porter, J. A., Jr., 1969. A survey for enteropathogenic bacteria in panamanian primates. *Lab. Anim. Care*, 19: 336-341.
6. Kourany, M., Myers, C. W., and Schneider, C. R., 1970. Panamanian amphibians and reptiles as carriers of *Salmonella*. *Am. J. Trop. Med. Hyg.*, 19: 632-638.
7. Kourany, M., and Vasquez, M. A., 1975. A survey to assess potential human disease hazards along proposed sea-level canal routes in Panamá and Colombia. VIII. Survey of enterobacterial pathogens in wild-caught vertebrates. *Milit. Med.*, 140: 22-25.
8. Edwards, P. R., and Ewing, W. H., 1962. *Identification of Enterobacteriaceae*. Burgess Publ. Co., Minneapolis, 250 pp.
9. Sickenga, F. N., 1964. Transmission of salmonellae and pathogenesis of salmonellosis in man. Pages 205-232 in Van Oye, E. (ed.), *The World Problems of Salmonellosis*. Dr. W. Junk Publ., The Hague.
10. Kourany, M., and Vasquez, M. A., 1969. Enteropathogenic bacteria associated with diarrhea among infants in Panamá. *Am. J. Trop. Med. Hyg.*, 18: 930-935.
11. Kourany, M., and Vasquez, M. A., 1974. A survey to assess potential human disease hazards along proposed sea-level canal routes in Panamá and Colombia. VI. Survey of enterobacterial pathogens in man. *Milit. Med.*, 139: 625-629.
12. Kourany, M., Vasquez, M. A., and Mata, L. J., 1971. Prevalence of pathogenic enteric bacteria in children of 31 Panamanian communities. *Am. J. Trop. Med. Hyg.*, 20: 608-615.